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OXOVANADIUM(IV) AND AMINO ACIDS—VIII. L-HISTIDINE DERIVATIVES AND RELATED LIGANDS: A SPECTROSCOPIC STUDY

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Abstract—The coordination structures of various species in aqueous solutions of oxovanadium(IV) + derivatives of L-histidine (1) and related ligands (3-methyl-L-histidine, 1methyl-L-histidine, L-histidine methyl ester, L-histidinol, $N\alpha$ -acetyl-L-histidine and histamine) have been studied by investigating the pH dependence of the circular dichroism, isotropic absorption and electron spin resonance spectra, and are compared with the structures of the corresponding complexes of L-histidine. Comparison of the spectroscopic results for all systems studied indicates coordination geometries.

Knowledge of the complex equilibria of VO^{2+} in the presence of amino acids is relevant in understanding its possible interaction with likely biological ligands. V—N(his) bonding has been inferred in some enzymes.¹ The design of model systems, preferably with L-histidine (L-his) as a ligand, and a better knowledge of the coordination modes of this amino acid to VO^{2+} are therefore of interest.

The crystal and molecular structure of $[(CH_3)_4N]$ $[VO^{IV}(L-hisO^-)(NCS)_2] \cdot H_2O$ has been determined by X-ray diffraction.² L-his functions as a tridentate chelate with the carboxylate group coordinating *trans* to the vanadyl bond, the V—N(imidazole) bond length (2.081 Å) being somewhat shorter than the V—N(amino) bond length (2.132 Å). The crystal and molecular structure of an oxo-bridged dinuclear V^{III} complex $[V_2^{III}(L-hisO^-)_4(\mu-O)] \cdot 2H_2O$ have also been reported.³ We recently reported^{4,5} studies on the L-histidine + VO²⁺ system in aqueous solution by combining the results of potentiometric and spectroscopic techniques. The present objective is to clarify the structures of species formed in this system. This was done by a comparison of the pH dependence of the visible circular dichroism, visible isotropic absorption and electron spin resonance (ESR) spectra obtained for the L-histidine (1) + VO²⁺ system with the corresponding spectra for several L-histidine derivatives and related ligands: 3-methyl-L-histidine (2), 1-methyl-L-histidine (3), L-histidine methyl ester (4), L-histidinol (5), N\alpha-acetyl-L-histidine (6) and histamine (7).

For the formulations $(VO)_p(ligand)_qH_r$, we normally use the abbreviation $M_pL_qH_r$. All species will be compared with the corresponding stoichiometries for the L-histidine + VO^{2+} system which are defined according to the reaction $(L^- = 1)$:

$$p\mathbf{M}^{2+} + q\mathbf{L}^{-} + r\mathbf{H}^{+} \Longrightarrow \mathbf{M}_{p}\mathbf{L}_{q}\mathbf{H}_{r}.$$
 (1)

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(1)







(7)

EXPERIMENTAL

All solutions were prepared in an inert atmosphere (high purity N_2 passed through soda lime and glass wool). All measurements were made at $25^{\circ}C$ with solutions containing 2.25 M NaNO₃. VO²⁺ and NaOH solutions were prepared and standardized as described in Part I.⁶ Other reagents are commercially available and were used without further purification.

Spectroscopic measurements

Visible spectra were recorded with a Perkin-Elmer lambda 9 spectrophotometer and circular dichroism with a Jasco 720 spectropolarimeter with a red sensitive photomultiplier. Cell compartments were kept at 25°C with circulating water from thermostatted baths. Unless otherwise stated, by visible (vis) and circular dichroism (CD) spectra we mean a representation of ε_m or $\Delta \varepsilon_m$ values vs λ $[\varepsilon_{\rm m} = {\rm absorption}/(b \times C_{\rm VO})$ and $\Delta \varepsilon_{\rm m} = {\rm differential}$ absorption/ $(b \times C_{vo})$; $b = optical path and C_{vo} =$ total oxovanadium(IV) concentration]. The Xband ESR spectra were normally recorded at 77 K (on glasses made on freezing solutions in liquid nitrogen) with a Bruker ESR-ER 200tt connected to a B-MN C5 ESR spectrometer and to a Bruker ESR data system (linked to an IBM AT computer). Spectra were normally recorded varying the pH with approximately fixed total vanadium and ligand concentration, at several ligand-to-metal (L/M)ratios.

TLC experiments

These were performed on Merck TLC plates (Art. 5626, 10×20 cm). The ligands and solutions used for spectroscopic measurements were monitored to check their purity and reactions (e.g. hydrolysis) that might possibly occur during the experiments. Normally, 2 μ l samples were applied to the plates 20 mm from the bottom. Elution was carried out in Camag twin chambers with walls covered with filter paper impregnated with the eluent. This was butanol-ethanol-propionic acidwater (10:10:2:5). When it reached \sim 120 mm from the bottom the plates were removed and dried. The chromatogram was developed with a ninhydrin-collidine (2,4,6-trimethylpyridine)-copper solution prepared according to Moffat and Lytle.⁷ In some cases, after development with this solution, the plate was placed in an enclosed chamber for development with iodine vapours. Typically samples were taken for TLC after dissolution of the ligand, and addition of VO^{2+} for several pH values.

RESULTS

3-Methyl-L-histidine + VO²⁺ system

Solutions of this amino acid gave only one reddish-brown spot in TLC experiments, and no decomposition of the ligand was detected throughout the measurements.

The ESR spectra with frozen "solutions" containing 3-methyl-L-histidine (3Mehis) and VO²⁺ at 77 K for high L/M ratios are like those obtained in the L-histidine + VO²⁺ system in similar experimental conditions throughout the whole pH range. The field region that corresponds to A_{\parallel} and $M_{\rm I} = 5/2$ and 7/2 (3800/4300 Gauss) gives more information about the type and number of species present. Figure 1 shows some of the results in this field range. The only noticeable difference between these spectra and those for L-his^{4,5} is that components IV are sharper and more symmetrical. Species IVA mentioned for the L-his + VO²⁺ system⁵ is either less important in the 3-Mehis + VO²⁺ system, or cannot be distinguished from V and VI (Fig. 1).

Table 1 summarizes the relevant g and A (\perp and \parallel) parameters calculated using the appropriate equations⁸ based on Chasteen's method.⁹ For most solutions at least two species are present and we have analysed these spectra as a superposition of two (or more) axial spectra. In some cases coincident perpendicular lines had to be assumed for the two (or more) species because these lines were not resolved in the spectra. In many cases even the parallel lines are close to each other, and the accuracy of the estimates of the spin Hamiltonian parameters is therefore affected. Despite this, the ESR parameters obtained for the 3Mehis+VO²⁺ system (Table 1) are almost identical to those of the corresponding species in the L-his+VO²⁺ system.^{4,5}

The visible and CD spectra obtained for solutions containing 2 and VO²⁺ are very similar to those for the L-his+VO²⁺ system,^{4,5} both in values of λ_{max} and the corresponding ε_m or $\Delta \varepsilon_m$. Consequently, we include no experimental spectra.

1-Methyl-L-histidine + VO²⁺ system

Solutions of this amino acid corresponded to bluish-brown spots in TLC experiments; a small yellowish-white spot is detected above the amino acid spot for solutions also containing oxovandium(IV). No decomposition of the ligand was detected during the measurements.

Figure 2A shows some of the ESR spectra obtained for "solutions" containing 1-methyl-Lhistidine (1Mehis) and VO²⁺ at 77 K. For $pH \leq 3.6$, the results are similar to those obtained for L-his and $3Mehis + VO^{2+}$ in similar conditions. However, for $pH \ge 3.7$, the ESR spectra differ markedly. IV and VI are not detected and in the pH range 6-9 a different species, designated by VIII', is dominant. At pH \sim 4.9 (Fig. 2), species designated by II', III' (and V') are dominant and the field positions correspond to II, III and V in the L-his^{4,5} and $3Mehis + VO^{2+}$ systems. In the experimental conditions of Fig. 2, for $pH \ge 8$ the solutions become brown and for pH \geq 9.5 oligometric species must form since the ESR signal almost disappears at pH \sim 11. Table 1 summarizes the relevant g and



Fig. 1. High field range (3800–4400 Gauss) of the first derivative ESR spectra at 77 K of frozen "solutions" containing 3-methyl-L-histidine and VO²⁺ with L/M = 9.63 and $C_{VO} \approx 0.011-0.012$ M. pH values and colours of the corresponding solutions are indicated.

A (\perp and \parallel) parameters calculated using the correct equations⁸ based on Chasteen's⁹ method.

The visible and CD spectra for solutions containing 3 and VO²⁺ are quite different from those in the 3Mehis and L-his+VO²⁺ systems in similar conditions.^{4,5} Notably, the λ_{max} (of the visible and CD spectra) differ and the $\Delta \varepsilon_m$ values are much lower (e.g. Figs 3 and 4).

$N\alpha$ -Acetyl-L-histidine

Solutions of this amino acid gave a single spot in TLC experiments ($R_f \sim 0.22$) after development with iodine vapour. No decomposition of the ligand was detected throughout the measurements.

Solutions containing Na-acetyl-L-histidine (Achis) and VO²⁺ (e.g. Figs 2B, 5 and 6) have a light blue colour till pH ≈ 4.5 . This distinguishes

them from those containing 3Mehis + VO²⁺ in similar conditions, which at pH \approx 3.8 are dark blue and for pH > 4.0–4.3 become violet. For pH \geq 4.8–5 the solutions become green (for the experimental conditions of Figs 2B, 5 and 6) and if the pH is not rapidly increased to 7–8, vanadyl hydroxide precipitates. For pH > 8, the solutions are greenish-brown but oxovanadium(IV) is extensively hydrolysed. The pH can then be decreased, the solutions become green, but the equilibria are sluggish and the visible and CD spectra recorded are irreproducible.

Figure 2B includes some of the ESR spectra. For $pH \ge 6$, the signal decreases and for $pH \ge 9$ no signal is detected [except for very high pH when $VO(OH)_3^-$ forms]. Table 1 includes ESR parameters calculated^{8,9} for the species detected.

Figure 5 includes visible spectra for solutions

containing	nine	g	1.930										1.945					
Table 1. Vanadium hyperfine coupling constants $A_1 \times 10^4$ /cm ⁻¹ and g_1 values estimated ^{8,9} from the first derivative ESR spectra at 77 K of frozen "solutions" oxovanadium(IV) and L-histidine or related ligands using high L/M ratios (see text)	Histar	۴	183	pH 1.7–3.9									~ 162	(XII [#])	,			
	L-Histidinol	g	1.929				~ 1.936						~ 1.952		~ 1.953			
		H	182	pH 1.7–3.8			1779	(1 *)					~ 163	(^ II ^)	158-160	(IX*)		
	ıethyl ester	g	1.932										~ 1.959		~ 1.949		~ 1.949	
	L-Histidine m	$oldsymbol{H}^{\parallel}$	183	pH 3.3									~ 162	(⊕VI)	~ 165	$(\Psi \oplus \simeq VI^{\oplus})$	~ 163	(⊕IIIA)
	Na-Acetyl-L-histidine	g _{II}	1.930		1.931		1.935		1.936						1.952			
		$oldsymbol{H}^{\parallel}$	183	pH 1.4	182	pH 2.0	178	pH 2.93	175	pH 4.34	168-173	(IX")			165	pH 7–8		
	1-Methyl-L-histidine	g_{\parallel}	1.936		1.940		1.938				~ 1.948		~ 1.948		~ 1.97			
		$oldsymbol{A}_{\parallel}$	181	pH 1.9	180	pH 2.46	178	pH 3.58			~ 170	pH 4.89	163-4	(,III,)	~ 150	(Imp)		
	3-Methyl-L-histidine	g	1.933		1.937		1.937		~ 1.947		~ 1.945		~ 1.957		~ 1.945		~ 1.953	
		\mathbf{A}^{\parallel}	182	pH 1.7	177	pH 2.65	177	pH 3.35	~ 172	E	~ 171	(III)	~ 161	(VI)	~ 166	E	~ 165	(VI)
	L-Histidine	g_	1.938		1.937		1.936		~ 1.945		~ 1.947		~ 1.953		~ 1.943		~ 1.946	
		"F	179	pH 1.9	177	pH 2.84	176	pH 3.36	~ 172	(II)	~ 170	(III)	160–162	(VI)	~ 166	E	~ 165	(VI)

Oxovanadium(IV) and amino acids-VIII







Fig. 3. CD spectra of solutions containing 1-methyl-L-histidine and VO²⁺ with L/M = 10.1 and $C_{\rm VO} \approx 0.012-0.011$ M. The pH corresponding to each spectrum is indicated. The ESR and visible spectra corresponding to some of these solutions are given in Figs 2A and 4, respectively. The representation of $\Delta \varepsilon_m$ as pH is varied is given in Fig. 14B.

containing 6 and VO^{2+} . These are quite different from those for the L-his, 3-Mehis and 1-Mehis + VO^{2+} systems, but are relatively similar to those for the glycylglycine and glycylglycylglycine + VO^{2+} systems¹⁰ in similar conditions. Figure 6 includes CD spectra for solutions containing 6 and VO^{2+} . These spectra also differ completely from those obtained for the L-his, 3Mehis and 1Mehis + VO^{2+} .

L-Histidine methyl ester

TLC chromatograms of solutions containing Lhistidine methyl ester (HisME) showed three spots: (1) due to L-his (violet; weak; $R_f \sim 0.04$), (2) due to 4 (brownish-violet; strong; $R_f \sim 0.18$), (3) due to an unknown compound (brownish-yellow; weak; $R_f \sim 0.27$). The addition of oxovanadium(IV) apparently does not change their relative intensities.

Fig. 4. Visible absorption spectra of solutions containing 1-methyl-L-histidine and VO^{2+} with L/M = 10.1 and $C_{VO} \approx 0.012-0.011$ M. The pH corresponding to each spectrum is indicated. The ESR and CD spectra corresponding to some of these solutions are given in Figs 2A and 3, respectively.

At pH ~ 10, the intensity of the L-his spot increases slowly with time and after ~2 h a new brownishyellow spot is detected at $R_f \sim 0.09$.

Solutions containing 4 and VO²⁺ with high L/M ratios, in experimental conditions similar to those of Figs 7–10, are light blue up to pH \approx 4. For pH > 4.1–4.3 (depending on experimental conditions), vanadyl hydroxide may precipitate if the pH is not rapidly increased to ~7.5. For pH > 8,

the solutions are grey-violet and for pH > 8.5-9they become brown, indicating extensive oxovanadium(IV) hydrolysis.

Figure 7A includes some of the ESR spectra. For pH < 3.5, the ESR parameters (Table 1) are consistent with the existence of $[VO(OH_2)_5]^{2+}$. For pH > 5-6, the present spectra show some similarities to those found for the L-his^{4.5} and 3Mehis + VO²⁺ systems, but the species designated

Fig. 5. Visible absorption spectra of solutions containing $N\alpha$ -acetyl-L-histidine and VO^{2+} with L/M = 15.0 and $C_{VO} \approx 0.012$ -0.011 M. The pH corresponding to each spectrum is indicated. The ESR and CD spectra corresponding to some of these solutions are given in Figs 2B and 6, respectively.

by IV^{\oplus} is much less important than for L-his and 3Mehis + VO^{2+} . The estimated ESR parameters are given in Table 1.

Figures 8 and 9 include some of the visible and CD spectra for solutions containing 4 and VO^{2+} . The pH was progressively increased from ~1.7 to 4.1, and spectra were recorded every 0.5 pH units. At pH ~ 4.1, after ~15 min vanadyl hydroxide began precipitating. Base was added till pH ~ 7.4, the very small amount of precipitate that had formed dissolved and the solution became greyish-violet. Spectra were recorded every 0.4–0.5 pH units till pH ~ 10, and then acid was added till pH ~ 6. Spectra were then recorded every 0.5 pH units till pH ~ 4.6.

For pH > 3, the CD (and visible) spectra differ from those for the L-his^{4,5} and 3Mehis + VO²⁺ systems in similar conditions. In particular, the maximum values of $\Delta \varepsilon_m$ (band I) in the pH range 5–7 are ~85% of the counterparts for solutions containing L-his or 3Mehis + VO²⁺. In the pH range 7–8, a band with $\Delta \varepsilon_m > 0$ is seen in the CD spectra of Fig. 9B. No such band is detected for the L-his^{4,5} and 3Mehis + VO²⁺ solutions.

L-histidinol

Solutions of this amino acid in TLC experiments showed a major violet spot at $R_f \sim 0.06$ and a yellowish-white spot at $R_f \sim 0.13$. No decomposition of the ligand was detected throughout the measurements.

Solutions containing 5 and VO²⁺ with high L/M ratios in conditions like those of Figs 7, 10 and 11 are light blue up to pH \approx 4, when they become dark blue. For pH > 4.0–4.2 (depending on conditions), vanadyl hydroxide may precipitate if the pH is not rapidly increased to ~8. For pH > 8, the solutions are light brown and the colour darkens for pH > 10, indicating extensive oxovanadium(IV) hydrolysis, also evident from the visible spectra (Fig. 10). On acidifying the solution corresponding to spectrum no. 16 in Fig. 11C, it becomes dark green at pH \approx 7.8. At pH \approx 7 vanadyl hydroxide precipitates.

Figure 7B includes some of the ESR spectra. For pH < 3.8, the estimated g and A (\perp and \parallel) parameters for the main species detected (Table 1) are consistent with the formation of $[VO(OH_2)_5]^{2+}$. For pH > 8, the spectra differ from those found for the L-his^{4,5}, 3Mehis and HisME+VO²⁺ systems. The species designated by IV* is much less important in the present system than IV in the L-his and 3Mehis+VO²⁺ systems (e.g. Fig. 1). The estimated^{8,9} ESR parameters are given in Table 1.

Although the CD spectra for $pH \leq 2.2$ (Fig. 11A) resemble those for the HisME + VO²⁺ system in similar conditions, most of the CD spectra are quite different from those for the other systems. In Fig. 11B, the pattern of the spectra clearly changes

Fig. 6. CD spectra of solutions containing $N\alpha$ -acetyl-L-histidine and VO^{2+} with L/M = 15.0 and $C_{VO} \approx 0.012-0.011$ M. The pH corresponding to each spectrum is indicated. The ESR and CD spectra corresponding to some of these solutions are given in Figs 2B and 5, respectively. Spectra 9–15 were recorded following this order, i.e. by successive addition of acid. As mentioned in the text the equilibria are sluggish, the spectra are not reproducible from batch to batch, but their general pattern is the same for each pH. The representation of $\Delta \varepsilon_m$ as pH is varied is given in Fig. 14A.

drastically as pH is increased from 8.3 to 9.9, an isodichroic point being detected at ~ 628 nm. Therefore, although no clear modifications are seen in the corresponding ESR (Fig. 7B) and visible (Fig. 10B) spectra, these CD spectra suggest the existence of an equilibrium between (at least) two optically active complexes. This involves VII* (and

IV*; VI*) and IX*. Besides the colours (and spectra) of the solutions, the relatively low $\Delta \varepsilon_m$ values indicate that vanadium is quite extensively hydrolysed in these conditions. For pH > 10, there is a general decrease in the $\Delta \varepsilon_m$ values as pH is increased, indicating further hydrolysis and gradual substitution of 5 by OH⁻.

Fig. 8. Visible absorption spectra of solutions containing L-histidine methyl ester and VO^{2+} with L/M = 20.0 and $C_{VO} \approx 0.015-0.008$ M (see text for details). The pH corresponding to each spectrum is indicated. The spectra are numbered in their order of recording. The ESR and CD spectra corresponding to some of these solutions are given in Figs 7A and 9, respectively.

Histamine

Solutions of 7 gave a single spot (brown or brownish-yellow, depending on the amount applied) in TLC experiments. No decomposition of the ligand was detected by TLC throughout the measurements.

Solutions containing 7 and VO²⁺ with high L/M ratios in conditions similar to those of Figs 12 and 13 have a light blue colour up to pH \approx 4. For pH > 4, vanadyl hydroxide precipitates if the pH is not rapidly increased above ~8. For pH > 8, the solutions are brown, indicating extensive oxovanadium(IV) hydrolysis, also evident from the visible spectra (Fig. 13). Acidifying the solution corresponding to spectrum no. 8 in Fig. 13, for pH < 8, vanadyl hydroxide precipitates.

Figure 12 includes some ESR spectra. For pH < 3.8, the estimated g and A (\perp and \parallel) parameters for the main species detected (Table 1) are consistent with the formation of $[VO(OH_2)_5]^{2+}$. For pH > 8, the spectra differ from those for the other systems in this study, and at pH 8.3 only one species is detected, designated by VII[#]. The ESR signal is much weaker than at lower pH, consistent with hydrolysis of oxovandium(IV), forming oligomeric species which are ESR silent.

Figure 13 includes some visible spectra of solutions containing 7 and VO²⁺. Up to pH ~ 3.5, the ε_m values are almost identical with those for $[VO(OH_2)_5]^{2+}$. At pH ~ 3.9, the general increase in the ε_m values is consistent with the hydrolysis and formation of $[(VO)_2(OH)_2]^{2+}$ (and $[VO(OH)]^+$).¹¹ For pH > 8, band II cannot be located and λ_{max} (band I) shifts to the red.

DISCUSSION

We have recently investigated^{4,5} the L-histidine + VO²⁺ system in aqueous solution by combining the results of potentiometric and spectroscopic techniques, and proposed an equilibrium model including species MLH₂, MLH, MLH₋₂, ML₂H₄, ML₂H₃, ML₂H₂, ML₂H, ML₂, ML₂H₋₁ and M₂L₂H₋₄. Isomeric structures for each stoichiometry were discussed, but it was important to clarify these for some species.

Several potentiometric and spectroscopic studies have been reported on the $Cu^{II} + L$ -his and related systems, particularly using CD techniques.¹¹⁻¹³ Most used L/M ratios of 2 and involved comparisons with binary and ternary systems. However, with VO^{2+} , studies involving solutions with L/M = 2 are possible only with very extensive hydrolysis of the metal ion : even when using high L/M ratios to prevent this, more than one stoichiometry is normally present at any pH value. Besides, due to the existence of the V=O bond, the number of possible isomeric structures is often

Fig. 9. CD spectra of solutions containing L-histidine methyl ester and VO^{2+} with L/M = 20.0 and $C_{\rm vo} \approx 0.015-0.008$ M (see text for details). The pH corresponding to each spectrum is indicated. The spectra are numbered in the order of recording; $\Delta \varepsilon_m$ values for spectra 1-6 are multiplied by a factor of 2. Some corresponding ESR and visible spectra are given in Figs 7A and 8, respectively. The representation of $\Delta \varepsilon_m$ as pH is varied is given in Fig. 14D.

greater than for Cu^{II} complexes.^{6,14} Consequently, what was done for the Cu^{II} systems ^{11–13} is not possible for vanadium: that would include the use of additive functions of independent contributions from groups present in the molecules or any detailed discussion about the origins of particular CD bands. However, comparisons between the pH dependence of the visible CD, visible isotropic absorption and ESR spectra obtained for the L-

histidine + VO²⁺ system and those for the corresponding spectra for several L-histidine derivatives and related ligands included in this work allow some clarification of the coordination modes. Representations of $\Delta \varepsilon_m$ values as pH is varied, such as those shown in Fig. 14, are also useful to detect the formation and disappearance of complex species in solution.

We now discuss, on the basis of the present ESR,

Fig. 10. Visible spectra of solutions containing L-histidinol and VO^{2+} with L/M = 15.3 and $C_{VO} \approx 0.018-0.015$ M (see text for details). The pH corresponding to each spectrum is indicated. Some corresponding ESR and visible spectra are given in Figs 7B and 11, respectively.

visible and CD spectra and those for L-his + oxovanadium(IV),^{4.5} structures for the complexes present in the several systems.

3-Methyl-L-histidine

The ESR, visible and CD spectra for solutions containing 2 and VO²⁺ closely resemble those for the L-his +VO²⁺ system,^{4,5} and the A_{\parallel} and g_{\parallel} parameters for the corresponding species in the two systems are almost identical (e.g. Table 1). Therefore, apart from small differences that may exist in the ratios of isomeric structures for particular stoichiometries, all results indicate that the mode of coordination of both ligands throughout the pH range up to at least 11–12 is the same. In particular, it may be concluded that no deprotonation/ coordination of the imidazole N(3)H (pyrrole NH) occurs (no comparative studies were done for pH > 12).

$N\alpha$ -Acetyl-L-histidine

The type of results obtained for this system (see above) and the fact that for $pH \ge 3.5$ the ESR, visible and CD spectra are quite different from those for the L-his and 3Mehis+VO²⁺ confirms

Fig. 11. CD spectra of solutions containing L-histidinol and VO²⁺ with L/M = 15.3 and C_{vo} ≈ 0.018 -0.015 M (see text for details). The pH corresponding to each spectrum is indicated. The ESR and visible spectra corresponding to some of these solutions are given in Figs 7B and 10, respectively. The representation of $\Delta \varepsilon_m$ as pH is varied is shown in Fig. 14C and E.

that the amino nitrogen (N_{am}) in the latter coordinates.

At pH ~ 1.4, the estimated g and A (\perp and \parallel) parameters for species IA" (Fig. 2B and Table 1) are consistent with the formation of $[VO(OH_2)_3]^{2+}$. For pH < 3 (e.g. Figs 2B, 5 and 6A) the results are consistent with the simultaneous existence of the aqua ion and of carboxylate complexes 8 (and 9 in lower concentration; Table 2). For pH > 3, the $\Delta \varepsilon_{\rm m}$ values start to increase (e.g. Figs 6A and 14A). At pH 3.5 they are still <0, but at pH 4.0 (spectrum no. 6) two bands with $\Delta \varepsilon_{\rm m} > 0$ are clearly seen, with $\lambda_{\rm max}$ (band I) ~775 and $\lambda_{\rm max}$ (band II) ~590 nm. A different species forms (II" in the ESR): this dominates the pattern of the CD spectra in the pH range 4.0-4.8. Assuming that the imidazole nitrogen (N_{im}) has a contribution to the A_{\parallel} values identical to the bipyridyl nitrogen atoms

Fig. 12. High field range (3800–4400 Gauss) of the first derivative ESR spectra at 77 K of frozen "solutions" containing histamine and VO²⁺ with L/M = 22.4 and $C_{\rm VO} \approx 0.018$ –0.014 M. Up to pH ~ 3.9, these were light blue; at pH ~8.3 the solution is brown. The corresponding wighle energy are shown in Fig. 12

responding visible spectra are shown in Fig. 13.

 $(162.8 \times 10^{-4} \text{ cm}^{-1})$,⁹ the ESR parameters for II" (Fig. 2B and Table 1) are consistent with the formation of a complex with two H₂O, one COO⁻ and N_{im} coordinated in equatorial positions. Structures 10 and 11 can then be envisaged for this species. In 10, the equatorial bidentate coordination of COO⁻ and N_{im} involves a seven-membered chelate ring, normally not very favourable. This may explain why for pH > 4.8 oxovanadium(IV) is very extensively hydrolysed and eventually vanadyl hydroxide precipitates.

One must also emphasize that structure 9 (and possibly 12) is also reasonable. In fact, the pattern of the CD spectrum for the corresponding species in the L-ala⁶ and L-ser¹⁵ + VO²⁺ systems has some similarities with that in the pH range 3–4.8 shown in Fig. 6A. Besides, the visible spectra for pH < 4.8 (e.g. Fig. 5) do not indicate equatorial coordination of a nitrogen atom. Dessi *et al.*¹⁶ also explain ESR spectra of solutions containing *N*-acetyl-glycine with L/M = 77 for pH < 6 assuming the formation of complexes such as 9 (Table 2).

At pH 5.95, species IX" is detected in the ESR, but a significant fraction of oxovandium(IV) is in oligomers. The ESR signal is weak and the accuracy of its parameters is correspondingly low. The ESRactive species could have coordination geometries such as 13 and 14. For pH > 7, the A_{\parallel} values of the ESR-active species decrease further. The $\Delta \varepsilon_m$ values continue to decrease and λ_{max} (bands I) shifts to the UV (Figs 6B and 14A). The absorbance for $\lambda < 550$ nm increases significantly and consequently the decreased accuracy of the CD spectra in this range precludes the determination of the true behaviour

Fig. 13. Visible absorption spectra of solutions containing histamine and VO²⁺ with L/M = 22.4and $C_{VO} \approx 0.018$ -0.015 M (see text for details). The pH corresponding to each spectrum is indicated. Some related ESR spectra are shown in Fig. 12. Spectra 1-5 and 6-8 have different ε_m scales.

of band II. These results are consistent with the progressive equatorial coordination of OH⁻ ligands till oxovanadium(IV) is completely hydrolysed.

l-Methyl-L-histidine

The results for this system are similar to those for L-ala + oxovanadium(IV),⁶ implying coordination as a simple α -amino acid. The spectra differ greatly from those for the L-his and 3Mehis + VO²⁺ systems. This further indicates coordination of N_{im} in those systems, where it is important in preventing oxovanadium(IV) hydrolysis as pH is increased.

At pH ~ 1.9, the estimated ESR parameters (Fig. 2A and Table 1) are consistent with the formation of $[VO(OH_2)_5]^{2+}$, the carboxylate complexes 8 probably existing in lower concentration. At pH 2.46 and 3.58 (Fig. 2A), the ESR spectra are consistent with the formation of the aqua ion and of carboxylate complexes 8 (and 9 in lower concentration). At pH 3.58, a second distinct species, designated by II', is detected. In the corresponding visible and CD spectra (spectra no. 4 in Figs 3A and 4A) band II shifts to the UV and $|\Delta \varepsilon_m|$ values increase significantly (Fig. 14B). These observations are consistent with the formation of 15 (and 16; Table 2).

At pH > 6, only one species is detected in the ESR spectra; this is designated by VIII' in Fig. 2A. The corresponding ESR parameters, visible and CD spectra are similar to those obtained for $[VO(L-ala)_2]$,⁶ except that the $|\Delta \varepsilon_m|$ values of band I are

about twice those for the bis-alaninato complex. Therefore, VIII' probably corresponds to structure 17; at higher values of pH structures such as 18 could also be important. For pH > 10, oligomeric species form: the ESR and CD signals disappear (Figs 2A, 3B and 14B) and the visible spectra change, as found for the other systems where oxovanadium(IV) hydrolyses extensively.

L-Histidinol

In the pH range 1.7–3.8, the estimated g and A(\perp and \parallel) parameters for the main species detected (Fig. 7B and Table 1) are consistent with the formation of [VO(OH₂)₅]²⁺. This agrees with the very low $\Delta \varepsilon_m$ values observed (Figs 11A and 14C, E) and with the visible spectra recorded (Fig. 10A). At pH 3.76, species I* is detected. The $|\Delta \varepsilon_m|$ values also increase significantly. Species I* could arise from an impurity, i.e. a small amount of histidine which at this pH would be in the form I (Fig. 1). As the R_f of 5 and 1 are similar, a small percentage of this ligand could hardly be detected by TLC. However, I* could also correspond to 19 whose estimated⁹ ESR parameters are approximately the same.

The CD spectra recorded in the pH range 3.5-4 suggest the formation of a very small amount of complexes involving the bidentate coordination of the ligand. At pH ~ 4, vanadium is very extensively hydrolysed and vanadyl hydroxide starts precipitating after several minutes. If base is added till pH ~ 8-9, the precipitate dissolves and the ESR and CD spectra become quite different (Figs 7B and

Fig. 14. Change with pH of the $\Delta \varepsilon_m$ values for solutions. (A) $N\alpha$ -Acetyl-L-histidine and VO^{2+} with L/M = 10.0 and $C_{VO} = 0.012-0.008$ M. (\blacksquare) 595 nm; (\triangle) 680 nm; (\square) 775 nm. (B) 1-Methyl-L-histidine and VO^{2+} with L/M = 15.0 and $C_{VO} = 0.012-0.011$ M. (\square) 535 nm; (\triangle) 590 nm; (\bullet) 590 nm; (\bullet) 650 nm; (\blacksquare) 740 nm. (C) L-Histidinol and VO^{2+} with L/M = 15.2 and $C_{VO} = 0.018-0.013$ M. (\blacksquare) 530 nm; (\triangle) 590 nm; (\square) 730 nm; (+) 830 nm. (D) L-Histidine methyl ester and VO^{2+} with L/M = 20.0 and $C_{VO} = 0.015-0.008$ M. (\blacksquare) 490 nm; (\triangle) 530 nm; (\square) 690; (*) 750 nm. (E) L-Histidinol and VO^{2+} with L/M = 15.2 and $C_{VO} \cong 0.018$ M. (\blacksquare) 530 nm; (\triangle) 590 nm; (\square) 730 nm; (+) 830 nm. Expanded scales of C for the low pH range.

11). However, oligomers are present in significant concentrations, as indicated by the brown colour of the solutions and the visible spectra (Fig. 10B).

Restricting our discussion to the monomeric species present, at pH ~ 8.4 the solution probably contains a mixture of VII* and IV*, possibly cor-

responding to structures 20 and 21, respectively (whether VI* and VII* are distinct species is uncertain). For pH ≥ 8 , the ESR signal (Fig. 7B) shifts to lower field values, but this may be due to the appearance of IV*. The pattern of the CD spectra changes as pH is increased from 8.3 to 9.9, an iso-

Table 2. Coordination geometries^a and corresponding A_{\parallel} estimated according to Chasteen's table,⁹ assuming the contribution of N_{im} is 162.8×10^{-4} cm⁻¹

^a For several of the structures included, other isomers are possible.

^bAs either N_{am} or N_{im} could be coordinated equatorial, they are not specified. A_{\parallel} was estimated assuming A_{\parallel} (N) = 161.5 × 10⁻⁴ cm⁻¹.

dichroic point being detected at ~628 nm. These CD spectra suggest the existence of an equilibrium between (at least) two optically active complexes: VII* (and VI* ?) and IX*. Species IX* is responsible for the positive $\Delta \varepsilon_m$ values in the range 450–620 nm at pH ~ 10. Its coordination geometry involves the equatorial coordination of the R—O⁻ group of one histidinol ligand. Likely coordination geometries are e.g. **22** and **23**.

For pH > 10, there is a general decrease in the $|\Delta \varepsilon_m|$ values as pH is increased (Figs 11C and 14C), indicating further hydrolysis and gradual substitution of 5 by OH⁻.

Histidine methyl ester

In the pH range 1.7–3.7, the estimated g and A(\perp and \parallel) parameters for the main species detected (Fig. 7A and Table 1) suggest the formation of [VO(OH₂)₅]²⁺, consistent with the very low $\Delta \varepsilon_m$ values (Figs 9A and 14D) and visible spectra (Fig. 8). The presence of a small amount of L-histidine in the starting material may account for the greater ε_m values in the range 650–800 nm than for the Lhistidinol or histamine + oxovanadium(IV) in similar conditions.

In the pH range 4.5–10, the spectra were recorded in the order specified in Figs 8 and 9. The high ε_m values in the visible spectra (Fig. 8) indicate that a small amount of oligomeric species is present throughout this range. In the pH range 4.5–5.8, the CD spectra are like those for similar solutions containing L-his or 3Mehis+oxovanadium(IV), but the $|\Delta \varepsilon_m|$ values are ~20% lower. Species IV^{\oplus} and V^{\oplus} could correspond to coordination geometries such as **21** and **24**, respectively. This would mean that their concentration ratio is not pH dependent. If this is correct the changes in the relative intensities of the corresponding ESR peaks are due to the formation of VI^{\oplus} and VII^{\oplus}, detected very close to V^{\oplus}.

In the pH range 6–9, the pattern of the CD spectra for band II changes (Figs 9B and 14D) and becomes quite different from those obtained for solutions containing L-his or 3Mehis+oxovanadium(IV) in similar conditions. This corresponds to the formation of species VI^{\oplus} detected in the ESR. For pH > 9, the pattern of the CD spectra changes again and becomes similar to those for similar solutions containing L-his or 3Mehis+oxovanadium(IV). This now corresponds to the formation of species VII^{\oplus} detected in the ESR.

Species VI^{\oplus} could correspond to an equatorial $(N_{im})_2(N_{am})_2$ donor set and an OH⁻ group axial. Species VII^{\oplus} could correspond to structures such as **20**. However, this would probably have A_{\parallel} values lower than for IV^{\oplus} ,⁹ contrary to observation. Structures such as **25** cannot be ruled out, but they would not explain the high $|\Delta \varepsilon_m|$ values for species ML_2H_{-1} that forms in the L-his and 3Mehis+ VO^{2+} system and possibly corresponds to VII^{\oplus} .

Histamine

Up to pH ~ 4 the estimated g and A (\perp and \parallel) parameters for the main species (Fig. 12 and Table 1) are consistent with the formation of $[VO(OH_2)_5]^{2+}$. Indeed, the ε_m values of the visible spectra in the pH range 1.6–3.5 (Fig. 13) practically coincide with an oxovanadium(IV) solution.

The ESR spectrum at pH 3.89 (Fig. 12) is very similar to that at pH 3.76 for the histidinol + VO²⁺ system (Fig. 7B). In particular, the species I[#] in Fig. 12 probably corresponds to the same coordination geometry as I^{*}. At pH ~ 3.9, the visible spectrum indicates partial hydrolysis of oxovanadium (IV) (i.e. $[(VO)_2(OH)_2]^{2+}$, which is ESR silent, forms in a relatively low concentration).

As with 5, vanadyl hydroxide precipitates for pH > 4. If base is (rapidly) added before a significant amount of solid forms, for pH > 8 it dissolves to a brown solution. The ESR spectra differ from those for the other systems in this study, and a single species (VII[#]) is detected at pH 8.3. The ESR signal is much weaker than for the low pH region; oxovanadium(IV) is significantly hydrolysed to oligomeric species which are ESR silent (e.g. {[(VO)₂(OH)₅]⁻}_n¹⁴). Species VII[#] possibly corresponds to structure 21; however, structures such as 20 are also plausible.

These results indicate that for L-his and $3Mehis + VO^{2+}$ systems the coordination of the R-COO⁻ group is important. Its absence, as with 5 or 7, produces completely different spectroscopic results, and vanadyl hydroxide precipitation occurs even with L/M ratios as high as 20. For histamine, $pK_a(N_{im}H) = 6.07$ and $pK_a(N_{am}H) = 9.79$, compared to 6.03 and 9.09, respectively, for L-his in the same medium (25°C; I = 0.1 M).¹⁷ Besides the absence of COO^- donor group, the increase of 0.7 units in $pK_a(N_{am}H)$ for 7 also gives rise to lower availability for its coordination to VO^{2+} . The opposite effect occurs for 4: its $pK_{a}(N_{am}H)$ is probably significantly lower than for L-his, and although VO^{2+} hydrolyses appreciably for pH > 4, vanadyl hydroxide precipitation can be avoided.

CONCLUSION

For most natural amino acids (from proteins), solutions with low L/M ratios contain much hydro-

lysed VO²⁺. For the three amino acids L-histidine, L-cysteine and aspartic acid, it is possible to avoid this extensive VO²⁺ hydrolysis. Of these, only for L-histidine has a VO²⁺ α -amino acid complex been characterized by X-ray diffraction.²

Comparisons between the pH dependence of the visible CD, visible isotropic absorption and ESR spectra obtained earlier for the L-histidine $+ VO^{2+}$ system⁵ with those for several L-histidine derivatives and related ligands included in this present work clarify the coordination modes of the ligand L-histidine.

Among the conclusions, the tridentate coordination of one of the L-histidine ligands in the pH range 6–8 is clearly proven. In fact :

- (a) Substitution on the amino nitrogen, as in Nαacetyl-L-histidine, gives rise to extensive VO²⁺ hydrolysis and eventually precipitation.
- (b) Blocking the imidazole nitrogen, as in 1methyl-L-histidine, produces rather similar results.
- (c) When the COO⁻ group is modified, as in L-histidine methyl ester, L-histidinol and (especially) histamine, extensive VO²⁺ hydrolysis occurs, but species containing the $(N_{im)2}$ $(N_{am})_2$ donor set are detected.
- (d) The results obtained for 3-methyl-L-histidine $+ VO^{2+}$ are almost identical to those for L-histidine.

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